**Protocol for *S. sclerotiorum* isolation**

**Preparing potato Dextrose agar media (39g/L)**

For making 1L:

1. In a 2L flask, add 500mL of double distilled H2O
2. Weigh out 39g of PDA and add it to the 2L flask
3. Add 1mL chloramphenicol
4. Add enough double distilled H2O to bring the volume to 1L
5. Autoclave on liquid cycle at 121°C for 30 minutes

For making 500mL:

1. In a 1L flask, add 250mL of double distilled H2O
2. Weigh out 19.5g of PDA and add it to the 1L flask
3. Add 500μL chloramphenicol
4. Add enough double distilled H2O to bring the volume to 500mL
5. Autoclave on liquid cycle at 121°C for 30 minutes

**Disinfecting sclerotia**

1. Prepare 1% sodium hypochlorite solution and pour into small 50mL beaker
   1. Add 20mL of double distilled H2O to 5mL of 5% sodium hypochlorite
2. Submerge sclerotia in solution for 1 minute
3. Rinse with sterile double distilled H2O for 30 seconds
4. Blot sclerotia on kim wipes

Adapted from Sanogo, S., and N. Puppala. 2007. Characterization of a Darkly Pigmented Mycelial Isolate of *Sclerotinia sclerotiorum* on Valencia Peanut in New Mexico. Plant Disease 91(9): 1077–1082. doi: [10.1094/PDIS-91-9-1077](https://doi.org/10.1094/PDIS-91-9-1077).